Reprinted from

INDUSTRIAL CROPS AND PRODUCTS

An International Journal

Industrial Crops and Products 14 (2001) 135-138

Enrichment of protein, starch, fat, and sterol ferulates from corn fiber by fine grinding and air classification*

Y. Victor Wu a.*, Robert A. Norton b

Fermentation Biochemistry, National Center for Agricultural Utilization Research, Agricultural Research Service,
 U.S. Department of Agriculture, 1815 N. University St., Peoria, IL 61604, USA
 Mycotoxin Research Unit, National Center for Agricultural Utilization Research, Agricultural Research Service,
 U.S. Department of Agriculture, 1815 N. University St., Peoria, IL 61604, USA

Received 21 August 2000; accepted 19 December 2000



INDUSTRIAL CROPS AND PRODUCTS

Aims and Scope

INDUSTRIAL CROPS AND PRODUCTS, AN INTERNATIONAL JOURNAL, publishes papers reporting the results of original research, short communications and critical reviews on all aspects of industrial crops and products.

This covers a wide range of aspects of cultivation, crop improvement, crop compounds, processing and integrated chain control, all focusing on the exploitation of agricultural crops for industrial use.

The scope of the journal covers a vast range of crops and research disciplines. Crops should contain significant renewable resources such as:

- fibres and fibre compounds;
- e carbohydrates;
- oils and fatty acids;
- waxes, resins, gums, rubber and other polymers;
- o proteins;
- essential oils for ink, lubricants, plastics, cosmetics;
- biologically active compounds for pharmaceuticals, herbicides and insecticides, and preservatives.

Examples of new or potential crops are agave, cassava, crambe, cuphea, elephant grass, fibre hemp, flax, guar, guayule, jojoba, kenaf, lesquerella, maize, meadowfoam, oil palm, peas, plantago, potato, pyrethrum, rape seed, safflower, soybean, Stokes aster, sugar beet, sunflower, veronia and wheat.

Papers within the above indicated frame-work will be accepted if they cover or integrate research on:

- agronomic production and modelling;
- breeding, genetics and biotechnology;
- post-harvest treatment and storage:
- (bio)process technology;
- (bio)chemistry;
- product testing, development and marketing;
- economics, and systems analysis and optimization.

Editors-in-Chief

For the Americas

Dr. F.S. Nakayama, USDA-ARS, US Water Conservation Lab., 4331 East Broadway, Phoenix, AZ 85040, USA

For the rest of the world

Dr E. de Jong, Agrotechnical Research Institute (ATO), P.O. Box 17, 6700 AA Wageningen, The Netherlands

Book Review Editor

D.T. Ray, Department of Plant Science, University of Arizona, Tucson, AZ 85721, USA

Editorial Advisory Board

T.P. Abbott, NCAUR-ARS-USDA, Peoria, IL, USA

P. Ahlgren, University of Karlstad, Karlstad, Sweden

M.G. Blase, University of Missouri-Columbia, Columbia, MO, USA

- C. Botti, Universidad de Chile, Santiago, Chile
- D. Byrom, ZENECA LifeScience Molecules, Cleveland, UK
- P. Colonna, INRA, Nantes, France
- K. Cornish, USDA-ARS, Albany, CA, USA
- J.T.P. Derksen, SHR Timber Research, Wageningen, The Netherlands
- S.A. Graham, Kent State University, Kent, OH, USA

- R.W. Kessler, Fachhochschule Reutlingen, Reutlingen, Germany
- B. Mattiasson, Dept. of Biotechnology, Lund, Sweden
- P.L. Milthorpe, Condobolin, NSW, Australia
- A. Noguchi, JIRCAS, Tsukaba, Ibaraki, Japan
- Y. Popineau, INRA, Nantes, France
- G. Röbbelen, Institut für Pflanzenbau und Pflanzenzüchtung, Göttingen, Germany
- S.F. Thames, Dept. of Polymer Science, Hattiesburg, MS, USA
- H. Tournois, ATO-DLO, Wageningen, The Netherlands
- G. Wegener, Institute for Wood Research, Munich, Germany

Publication information: Industrial Crops and Product (ISSN 0926-6690). For 2001, volumes 13-14 are scheduled for publication. Subscription prices are available upon request from the Publisher or from the Regional Sales Office nearest you or from this journal's website (http://www.elsevier.nl/locate/indcrop). Further information is available on this journal and other Elsevier Science products through Elsevier's website: (http://www.elsevier.nl). Subscriptions are accepted on a prepaid basis only and are entered on a calendar year basis. Issues are sent by standard mail (surface within Europe, air delivery outside Europe). Priority rates are available upon request. Claims for missing issues should be made within six months of the date of dispatch.

The paper used in this publication meets the requirements of ANSI/NISO Z39.48-1992 (Permanence of Paper)



Industrial Crops and Products 14 (2001) 135-138

INDUSTRIAL CROPS AND PRODUCTS

AN INTERNATIONAL JOURNAL

www.elsevier.com/locate/indcrop

Enrichment of protein, starch, fat, and sterol ferulates from corn fiber by fine grinding and air classification*

Y. Victor Wu a.*, Robert A. Norton b

- * Fermentation Biochemistry, National Center for Agricultural Utilization Research, Agricultural Research Service, U.S. Department of Agriculture, 1815 N. University St., Peoria, IL 61604, USA
- ^b Mycotoxin Research Unit, National Center for Agricultural Utilization Research, Agricultural Research Service, U.S. Department of Agriculture, 1815 N. University St., Peoria, IL 61604, USA

Received 21 August 2000; accepted 19 December 2000

Abstract

Corn fiber contains hemicellulose, starch, cellulose, protein and fat. There is commercial interest to produce sterol ferulates from corn fiber oil, which accounts for only 3% of corn fiber by weight. An inexpensive process to enrich starch, protein, and fat contents of corn fiber is desirable for increased utilization and further processing. This study was conducted to determine whether fine grinding and air classification of corn fiber into separate fractions according to particle size could enrich selected components of interest. Corn fiber was finely ground in a pin mill at high speed, and the resulting ground fiber was separated into various fractions with cutpoints of 15, 18, 24 and 30 μ m. The finest fraction, with a particle size of less than 15 μ m, showed enriched protein, starch, fat and sterol ferulates contents compared with the starting corn fiber. The greater than 30 μ m fraction was separated by sieves into five fractions. In general, protein, starch, fat and sterol ferulates contents of the fractions decreased and total dietary fiber increased with increasing particle size. Fine grinding and air classification of corn fiber can enrich protein, starch, fat and sterol ferulates contents in the fine fractions and can make further processing more economical. Published by Elsevier Science B.V.

Keywords: corn fiber; protein; starch; fat; sterol ferulates; air classification

E-mail address: wuyv@mail.ncaur.usda.gov (Y.V. Wu).

1. Introduction

There is commercial interest to produce sterol ferulates from corn fiber oil; however, current processes are expensive because the oil content of corn fiber is only 3% by weight. An inexpensive process that can enrich valuable components in corn fiber is desirable for increased utilization.

Wet milling of corn separates the kernel into starch, germ, protein, steepwater, and fiber. Corn

0926-6690/01/S - see front matter. Published by Elsevier Science B.V.

PII: S0926-6690(01)00076-0

Names are necessary to report factually on available data; however, the USDA neither guarantees nor warrants the standard of the product, and the use of the name by USDA implies no approval of the product to the exclusion of others that may also be suitable.

^{*} Corresponding author. Tel: +1-309-6816364; fax: +1-309-6816686.

fiber contains 20-22% starch, 12-14% cellulose, 11-12% protein, 3-4% fat, and 35-40% hemicellulose (Carlson 1994, Saha et al., 1998). By treating corn fiber with enzyme, Leathers and Gupta (1996) reported that up to 70% of polymeric xylose and 100% of polymeric arabinose from corn fiber hemicellulose were liberated and most or all of the starch was hydrolyzed to glucose. Doner and Hicks (1997) isolated water-soluble hemicellulose B and waterinsoluble hemicellulose A by alkaline hydrogen peroxide extraction of corn fiber. Moreau et al. (1996) extracted corn fiber to yield an oil containing sterol ferulates, which may lower serum cholesterol levels (Moreau et al., 1998). Dowd (1997) recovered approximately one-half of the starch in corn fiber by a laboratory scale wet process involving grinding, screening, washing, and centrifuging. Wet processing is usually expensive because of the need for drying and wash water disposal. Thus, we studied the effect of fine grinding and air classification on enrichment of protein, starch, fat, and sterol ferulates in corn fiber.

2. Materials and methods

Wet, de-watered corn fiber from the wet milling of corn was supplied by Williams Energy Services (formerly Pekin Energy Co.), Pekin, IL. The wet corn fiber was dried overnight at 63°C in a Proctor Dryer (Procter & Schwartz, Horsham, PA). The dried corn fiber was ground in a hammer mill equipped with a 2.36 mm diameter opening screen.

The corn fiber was ground three times at 14,000 rpm in an Alpine model 160Z pin mill (Augsburg, Germany), and fractionated in a Pillsbury laboratory model air classifier (Minneapolis, MN). The classifier was first set at a 15 μ m cutpoint to obtain a fine and a coarse fraction. The coarse fraction was then classified successively with 18, 24 and 30 μ m cutpoints to obtain four fine fractions (1 through 4 in increasing size) and a coarse residue (fraction 5). The > 30 μ m coarse residue (fraction 5) from air classification was further separated by sieving successively by sieves with square openings of 89, 124, 147, and 175 μ m to obtain 30–89, 89–124, 124–147, 147–175, and > 175 μ m fractions. Pin milling, air classification and sieving experiments

were carried out in duplicate from the same batch of oven-dried corn fiber.

Nitrogen, moisture and ash were determined by the American Association of Cereal Chemists (AACC) Methods 46-16, 44-15A and 08-16, respectively (AACC, 1995). Nitrogen was determined by Kjeldahl, moisture was determined from weight loss at 130°C for 1 h, and ash was determined gravimetrically after heating at 600°C for 2 h. Protein was calculated by nitrogen \times 6.25. Starch was measured by polarimetry from a calcium chloride solution containing stannic chloride (Earle & Milner, 1944). This method is slightly different from the Corn Refiners Association (CRA) Method A-20 (CRA, 1986). Fat was measured by petroleum ether extraction in a Tecator Soxtec System 1043 extraction unit by the Association of Official Analytical Chemists (AOAC) Official Method 920.39, and total dietary fiber was determined by AOAC Official Method 985.29 (A-G) (AOAC, 1998). Composition studies were averaged from the duplicate samples.

For detecting sterols, a 0.5 g aliquot of sample was added to a 15×125 mm test-tube, 5 ml hexane added and extracted twice by mixing on a Vortex Genie 2 (Fisher Scientific, Pittsburgh, PA) at maximum speed for 60 min. Following each extraction, the tubes were centrifuged for 10 min at 3500 rpm to pellet the sample, and the supernatant was pipetted into a tared test-tube. Hexane was evaporated with a stream of nitrogen and gentle heat until the sample was dry, and the residue weighed.

The hexane extract was analyzed for sterol ferulates by high performance liquid chromatography (HPLC) using a diol column (5 μ m, 4.6 mm \times 250 mm, Lichchrosorb Diol; Alltech Associates, Inc., Deerfield, IL) and eluted with hexane:2-propanol (98:2) at a flow rate of 1.5 ml/min. The extract was dissolved in 2 ml of hexane and 20 µl was injected for each analysis. The effluent was monitored at 210 nm and 325 nm using a diode-array detector (Beckman, Model 168, Fullerton, CA). Samples were quantitated from peak heights and a standard curve was derived by linear regression using γ-oryzanol (CTC Organics, Atlanta, GA) and at 325 nm. Each sample absorbance was analyzed twice, and the results were averaged. Injections were made by autoiniector (Beckman, model 507). Under these HPLC conditions, the sterol ferulates eluted as a group at about 9.9 min. To check for coelution of other UV-absorbing compounds, which might bias quantitation, the UV spectrum of the top of each ferulate peak was retrieved and superimposed on the peak spectrum of γ -oryzanol. Examination of the spectra showed no appreciable differences. In addition, the UV spectrum at one-third peak height before and after the maximum were retrieved and superimposed on the equivalent spectrum from the standard, and again no appreciable differences were found between the samples as compared to the y-oryzanol standard. Duplicate samples were extracted by hexane and sterol ferulates determined by HPLC.

The data were treated by analysis of variance, and Tukey's Studentized range test was used to determine significant difference at P < 0.05 (SAS Institute, Cary, NC).

3. Results and discussion

The yield and composition of corn fiber and its air classified fractions after fine grinding are listed in Table 1. Fraction 1, the finest fraction with

particle size less than 15 µm, had higher protein. starch, fat, and sterol ferulates, but smaller total dietary fiber content, than the starting corn fiber. Fraction 2 has double the starch, higher fat and sterol ferulates, but lower total dietary fiber content than the starting corn fiber. Fraction 3 has higher starch, fat, and sterol ferulates than the corn fiber. Fraction 5 has lower protein, starch, fat, and sterol ferulates content than corn fiber. When fraction 5 was separated by sieves into different particle sizes, the protein, starch, fat, and sterol ferulates decreased, but total dietary fiber content increased, with increasing particle size. Ash contents of all fractions in Table 1 were not significantly different (P > 0.05) and averaged 0.5%, and therefore were not listed individually in the table.

Because the sterol ferulates and total dietary fiber concentrations are inversely correlated, the sterol ferulates do not appear to be associated with the bulk of the fiber. Seitz (1989) dissected tissues from corn and found that the sterol ferulates were associated mostly with inner pericarp, and the aleurone was in the inner pericarp fraction. Our results seem to agree with those of Seitz (1989). Because the aleurone layer is juxtaposed to the peripheral part of the endosperm, the highest concentrations of sterol ferulates are

Table 1 Yield and composition of air classified and sieved corn fiber fractions, % dry basis

Fraction	μm	Yield	Protein	Starch	Fat	TDF ^a	Sterol ferulates	Sterol ferrulates in oil
Corn fiber			13.1 CD	15.4 CD	2.7 DE	76.4 A	0.181 E	6.7 B
1	< 15	7.9 ^b CD	17.1 A	23.3 B	5.5 A	41.3 C	0.306 A	5.6 C
2	15-18	10.4 CD	13.5 BCD	30.2 A	4.0 B	45.4 BC	0.210 C	5.2 D
3	18-24	30.7 B	14.3 BC	23.0 B	3.1 C	59.8 ABC	0.224 B	7.2 A
4	24-30	9.2 CD	14.8 BC	13.8 DE	2.9 CDE	62.6 ABC	0.194 D	6.7 B
5	> 30	41.9 A	10.7 E	10.8 EF	1.8 F	80.6 A	0.100 G	5.5 C
5	30-89	10.7 CD	15.4 AB	19.9 BC	3.0 CD	58.1 ABC	0.112 F	3.7 F
5	89-124	12.3 C	11.7 DE	10.4 EF	2.5 E	72.2 AB	0.108 F	4.3 E
5	124-147	5.6 D	8.6 F	6.8 FG	1.9 F	79.5 A	0.074 H	3.9 F
5	147-175	6.7 CD	7.0 F	4.7 G	1.5 FG	83.0 A	0.064 I	4.3 E
5	>175	6.6 CD	6.9 F	4.0 G	1.2 G	79.4 A	0.050 J	4.1 E

[&]quot; Total dietary fiber.

b Values followed by the same length a column at an occasion of the contents were not significantly different (P>0.05) and averaged 0.5% 101 1010 1020

found in the fractions with the highest concentrations of starch and protein.

Moreau et al. (1996, 1999) found that both corn bran from dry milling and corn fiber from wet milling contain ferulate phytosterol esters. They indicated that corn bran oil contains up to 1.5% ferulate phytosterol esters, and that corn fiber oil may contain up to 6.75% ferulate phytosterol esters. Also, Moreau et al. (1998) observed that during dry milling most of the ferulate esters (about 80%) separate with the grits, but during wet milling they separate with the fiber. For comparison purposes, percent sterol ferulates in oil ranged from 3.7 to 7.2% for air classified and sieved fraction, and from 5.2 to 7.2% for the five air classified fractions (Table 1). No definite trend was present between particle size and percent sterol ferulates in oil. Although the 18-24 µm fraction contained higher sterol ferulates and higher percent sterol ferulates in oil (P < 0.05) than the whole fiber, the practical significance of this difference remains to be further evaluated. Taylor and King (2000) reported enrichment of ferulate phytosterol esters from corn fiber oil using supercritical fluid extraction and chromatography. Because classification is relatively inexpensive, our enriched fraction(s) can be further processed by supercritical fluid extraction of ferulate phytosterol esters or by other suitable processes.

4. Conclusion

Fine grinding and air classification of corn fiber can enrich protein, starch, fat and sterol ferulates contents in the fine fractions. Selective combination of the fractions can make further processing more economical for the component(s) of interest.

Acknowledgements

The authors thank B. D. Deadmond and A. M. Kelly-Webb for technical assistance.

References

- American Association of Cereal Chemists, 1995. Approved methods of the AACC, 9th ed. St Paul, MN.
- Cunniff, P. (Ed.), 1998. Official Methods of Analysis of AOAC International, 16th ed., 4th rev. Association of Official Analytical Chemists.
- Carlson, T., 1994. Industrial utilization of corn starch and corn fiber for increased ethanol yield: Summary Paper. Proc. Corn Utilization Conf. V. National Corn Growers Association, St. Louis, MO.
- Corn Refiners Association, 1986. Analytical Methods of the Member Companies of the Corn Refiners Association, Inc. Washington, DC.
- Doner, L.W., Hicks, K.B., 1997. Isolation of hemicellulose from corn fiber by alkaline hydrogen peroxide extraction. Cereal Chem. 74, 176–181.
- Dowd, M.K., 1997. Recovery of starch and protein from wet-milled corn fiber. Cereal Chem. 74, 589-593.
- Earle, F.R., Milner, R.T., 1944. Improvements in the determination of starch in corn and wheat. Cereal Chem. 21, 567-575.
- Leathers, T.D., Gupta, S.C., 1996. Saccharification of corn fiber using enzymes from Aureobasidium sp. strain NRRL Y-2311-1. Appl. Biochem. Biotechnol. 59, 337-347.
- Moreau, R.A., Hicks, K.B., Nicolosi, R.J., Norton, R.A., 1998.
 Corn fiber oil its preparation and use. US Patent 5 843, 499.
- Moreau, R.A., Powell, M.J., Hicks, K.B., 1996. Extraction and quantitative analysis of oil from commercial corn fiber. J. Agric. Food Chem. 44, 2149–2154.
- Moreau, R.A., Singh, V., Eckhoff, S.R., Powell, M.J., Hicks, K.B., Norton, R.A., 1999. Comparison of yield and composition of oil extracted from corn fiber and corn bran. Cereal Chem. 76, 449–451.
- Saha, B.C., Dien, B.S., Bothast, R.J., 1998. Fuel ethanol production from corn fiber. Current status and technical prospects. Appl. Biochem. Biotechnol. 70-72, 115-125.
- Seitz, L.M., 1989. Stanol and sterol esters of ferulic and p-courmaric acids in wheat, corn, rye, and triticale. J. Agric. Food Chem. 37, 662–667.
- Taylor, S.L., King, J.W., 2000. Enrichment of ferulate phytosterol esters from corn fiber oil using supercritical fluid extraction and chromatography. J. Am. Oil Chem. Soc. 77, 687-688.

Supplied by U.S. Dept. of Agriculture

Sational Center for Agricultural

Hillipation Research, Peoria, Illinois